

SUPPORTING INFORMATION

Synthesis of *o*-Nitrophenylphenyliodonium fluoride (NPIF, 1):

(a) *o*-Nitrophenylphenyliodonium iodide-To a stirred solution of *o*-iodonitrobenzene (5.0 g, 20 mmol) in H₂SO₄ (100 mL) was added K₂S₂O₈ (6.0 g, 22 mmol) in small portions followed by benzene (25 mL) at room temperature, and the mixture was stirred vigorously for 1.5 h. The reaction mixture was poured into ice (total volume 300 mL), and the insoluble material was removed by filtration. The filtrate was treated with aqueous KI (5 g/20 mL of H₂O), giving an orange precipitate that was filtered and washed thoroughly with H₂O (250 mL) followed by a small amount of acetone. The collected solid was dried over P₂O₅ under reduced pressure to give 7.95 g (87%) of *o*-nitrophenylphenyl-iodonium iodide. This procedure was developed from the method of Beringer et al. (ref 13a in the text of the paper. Note the typographical error in their original procedure: *o*-iodosonitrobenzene should be replaced with *o*-iodonitrobenzene). (b) *o*-Nitrophenylphenyliodonium fluoride-To a vigorously stirred solution of AgF (1.0 g, 7.9 mmol) in H₂O (30 mL) was added the iodonium iodide (3.6 g, 7.9 mmol) and the mixture was stirred for several hours. After removal of the insoluble material by filtration, the solution was concentrated below 30 °C under reduced pressure (< 5 torr). The residue was diluted with 8 mL of MeCN and crystallized at 0 °C to give NPIF (1, 1.80 g first crop, 0.33 g second crop; total 2.13 g, 78%).

1: ¹H NMR (DMSO-d₆) δ: 7.54 (2H, t, J = 7.5 Hz), 7.66-7.78 (4H, m), 7.13 (2H, d, J = 7.5 Hz), 8.29 (1H, d, J = 7.5 Hz). ¹³C NMR (DMSO-d₆) δ: 116.2, 120.0, 126.2, 131.6, 131.7, 131.8, 135.0, 136.2, 136.3, 148.0.

General Procedure for *o*-Nitrophenylation Reaction:

General procedure for arylation reaction: To a stirred solution of NPIF 1 (345 mg, 1 mmol) in dry DMSO-CH₂Cl₂ (1.4 mL-2.1 mL) was added a TMS ether 2 (1 mmol) dropwise at ca. -40 °C (CO₂-MeCN) under nitrogen. The mixture was stirred for 2 h at this temperature, and allowed to warm up to room temperature gradually over 2-3 h. The reaction mixture was poured into H₂O (10

mL), and the whole was extracted with ether (10 mL x 3). The extracts were washed with brine, dried over MgSO₄, and concentrated. The residue was purified by flash column chromatography on silica gel (EtOAc:hexane = 1:30 – 1:5) to give the aryl ketone (**3**). Aryl ketones **3** were characterized from ¹H, ¹³C NMR, and IR spectra. See supporting information for the spectral data as well as copies of actual spectra.

3a: ¹H NMR (CDCl₃) δ: 1.75-1.89 (2H, m), 2.01-2.12 (2H, m), 2.15-2.18 (1H, m), 2.33-2.38 (1H, m), 2.50-2.53 (2H, m), 4.28 (1H, dd, *J* = 5.5, 12.5 Hz), 7.34 (1H, d, *J* = 8 Hz), 7.39 (1H, t, *J* = 8 Hz), 7.57 (1H, t, *J* = 8 Hz), 7.95 (1H, d, *J* = 8 Hz). ¹³C NMR (CDCl₃) δ: 25.2, 26.9, 33.4, 41.9, 53.2, 124.6, 127.5, 130.1, 133.0, 133.8, 149.0, 207.9.

3b: Major isomer; ¹H NMR (CDCl₃) δ: 1.36 (3H, d, *J* = 7), 1.86-1.90 (2H, m), 2.01-2.12 (3H, m), 2.30-2.39 (1H, m), 2.67-2.75 (1H, m), 4.52 (1H, dd, *J* = 5, 13 Hz), 7.33 (1H, d, *J* = 8 Hz), 7.40 (1H, dt, *J* = 1, 8 Hz), 7.58 (1H, dt, *J* = 1, 8 Hz), 7.97 (1H, dd, *J* = 1, 8 Hz). ¹³C NMR (CDCl₃) δ: 17.0, 20.3, 32.7, 33.0, 44.8, 49.2, 124.9, 127.7, 130.1, 133.1, 134.2, 149.3, 211.8. Assignable peaks of minor isomer; ¹H NMR (CDCl₃) δ: 1.07 (3H, d, *J* = 6.5), 4.35 (1H, dd, *J* = 5, 13 Hz). ¹³C NMR (CDCl₃) δ: 14.6, 25.5, 34.2, 36.7, 45.8, 53.1, 124.8, 129.9, 133.0, 210.0.

3c: ¹H NMR (CDCl₃) δ: 0.87 (3H, d, *J* = 6), 1.60 (1H, dq, *J* = 3, 13 Hz), 1.87 (1H, tq, *J* = 4, 13.5 Hz), 2.03-2.06 (1H, m), 2.11-2.15 (1H, m), 2.26-2.31 (1H, m), 2.47 (1H, dt, *J* = 6, 14 Hz), 2.55-2.58 (1H, m), 3.82 (1H, br s), 7.28 (1H, d, *J* = 8 Hz), 7.40 (1H, t, *J* = 18 Hz), 7.58 (1H, t, *J* = 8 Hz), 7.92 (1H, d, *J* = 8 Hz). ¹³C NMR (CDCl₃) δ: 20.9, 25.2, 34.1, 39.4, 41.3, 60.7, 124.7, 127.7, 131.8, 132.6, 132.8, 150.1, 207.8.

3d: Major isomer; ¹H NMR (CDCl₃) δ: 1.10 (3H, d, *J* = 6.5 Hz), 1.54-1.63 (1H, m), 1.84 (1H, br q, *J* = 12.5 Hz), 2.11-2.19 (2H, m), 2.28-2.33 (1H, m), 2.50-2.55 (1H, m), 2.56 (1H, br dt, *J* = 6, 14 Hz), 4.36 (1H, dd, *J* = 5, 13 Hz), 7.35 (1H, d, *J* = 8 Hz), 7.42 (1H, t, *J* = 8 Hz), 7.60 (1H, t, *J* = 8 Hz), 7.99 (1H, dd, *J* = 1, 8 Hz). ¹³C NMR (CDCl₃) δ: 21.3, 32.2, 35.1, 41.3, 41.7, 52.4, 124.8, 127.8, 130.3, 133.1, 133.9, 149.2, 208.3. Assignable peaks of minor isomer; ¹H NMR (CDCl₃) δ: 1.32 (3H, d, *J* = 7), 1.92-1.95 (1H, m), 2.40-2.45 (1H, m), 2.26-

2.72 (1H, m), 4.45 (1H, dd, $J = 5.5, 12.5$ Hz). ^{13}C NMR (CDCl_3) δ : 17.6, 27.2, 32.1, 37.5, 39.2, 48.2, 130.6.

3e: Less polar isomer; ^1H NMR (CDCl_3) δ : 1.74-1.84 (2H, m), 2.03 (1H, dq, $J = 3, 12.5$ Hz), 2.13-2.22 (1H, m), 2.27-2.34 (2H, m), 3.43 (3H, s), 3.66 (1H, br t, $J = 3$ Hz), 4.85 (1H, dd, $J = 5, 12.5$ Hz), 7.39-7.45 (2H, m), 7.59 (1H, dt, $J = 1, 8$ Hz), 7.91 (1H, dd, $J = 1, 8$ Hz). ^{13}C NMR (CDCl_3) δ : 19.9, 33.8, 34.4, 48.4, 57.7, 84.1, 124.5, 127.8, 130.3, 132.8, 133.0, 149.6, 208.9. More polar isomer; ^1H NMR (CDCl_3) δ : 1.73 (1H, dq, $J = 3, 12.5$ Hz), 1.90-20.5 (2H, m), 2.08-2.12 (1H, m), 2.34-2.38 (1H, m), 2.46-2.51 (1H, m), 3.47 (3H, s), 4.11 (1H, dd, $J = 6, 11.5$ Hz), 4.40 (1H, dd, $J = 5, 13$ Hz), 7.39 (1H, d, $J = 7$ Hz), 7.43 (1H, t, $J = 7.5$ Hz), 7.62 (1H, t, $J = 7.5$ Hz), 8.03 (1H, dd, $J = 1, 8$ Hz). ^{13}C NMR (CDCl_3) δ : 23.4, 33.3, 35.1, 51.7, 58.0, 84.6, 125.0, 127.9, 129.6, 132.9, 133.3, 148.7, 206.5.

3f: ^1H NMR (CDCl_3) δ : 1.93-2.02 (1H, m), 2.13-2.22 (2H, m), 2.33-2.40 (1H, m), 2.44-2.56 (2H, m), 3.89 (1H, dd, $J = 8.5, 11.5$ Hz), 7.24 (1H, d, $J = 8$ Hz), 7.38 (1H, dt, $J = 1, 7.5$ Hz), 7.53 (1H, dt, $J = 1, 7.5$ Hz), 7.95 (1H, dd, $J = 1, 8$ Hz). ^{13}C NMR (CDCl_3) δ : 20.8, 31.2, 37.8, 53.8, 125.2, 127.9, 131.4, 133.3, 133.4, 149.1, 215.4.

3g: Major isomer; ^1H NMR (CDCl_3) δ : 1.18 (3H, d, $J = 6.5$ Hz), 1.54-1.62 (1H, m), 2.03-2.10 (1H, m), 2.33-2.41 (2H, m), 2.46-2.51 (1H, m), 3.84 (1H, dd, $J = 8.5, 12.5$ Hz), 7.22 (1H, d, $J = 8$ Hz), 7.38 (1H, t, $J = 8$ Hz), 7.54 (1H, t, $J = 7.5$ Hz), 7.94 (1H, d, $J = 8$ Hz). ^{13}C NMR (CDCl_3) δ : 14.7, 29.7, 30.1, 44.3, 53.3, 125.2, 127.9, 131.3, 133.3, 133.9, 149.2, 217.2. Assignable peaks of minor isomer; ^1H NMR (CDCl_3) δ : 1.56-1.67 (1H, m), 1.78-1.83 (1H, m), 2.10-2.17 (1H, m), 2.24-2.30 (1H, m), 2.49-2.56 (1H, m), 4.16 (1H, br t, $J = 10$ Hz), 7.17 (1H, d, $J = 8$ Hz). ^{13}C NMR (CDCl_3) δ : 15.3, 28.6, 41.6, 52.0, 125.0, 127.7, 130.4, 133.1, 217.6.

General Procedure for TiCl_3 Reduction:

Aqueous TiCl_3 can be purchased from Aldrich. Alternatively, it can be prepared inexpensively by dissolving $\text{TiCl}_3 \bullet 1/3 \text{ AlCl}_3$ (available from Strem Chemicals, Inc.) in water (CAUTION! Strong

exotherm!), filtering off the precipitate. The concentration of the purple solution can be determined by titrating with 0.1 M KBrO₃, wherein disappearance of the purple color indicates the endpoint. To a purple solution of 1.44 M TiCl₃ in H₂O (0.56 mL, 0.8 mmol of TiCl₃) was added 2.5 M NH₄OAc in H₂O (1 mL) followed by acetone (1 mL) at room temperature. The purple color turned dark brown, and the mixture formed a two-phase system. A solution of a nitro ketone **3** (0.1 mmol) in acetone (1 mL) was added dropwise at room temperature with vigorous stirring. After 15 min, the reaction mixture was diluted with H₂O (10 mL), and extracted with EtOAc (10 mL × 3). The extracts were washed with NaHCO₃ aqueous solution followed by NaCl aqueous solution, then dried over Na₂SO₄ and concentrated. The residue was purified by flash column chromatography on silica gel (EtOAc:hexane = 1:60 – 1:50, containing a small amount of Et₃N) to give the indole product (**4**). Indoles **4a**,² **b**,³ **d**,¹ **e**,⁴ **f**,² **g**³ were identified with reported ¹H NMR spectra. The known indole **4c**¹ was characterized from ¹H spectrum.

4c: ¹H NMR (CDCl₃) δ: 1.37 (3H, d, *J* = 7 Hz), 1.55-1.59 (1H, m), 1.79-1.83 (1H, m), 1.95-2.00 (2H, m), 2.69-2.71 (2H, m), 3.11-3.13 (1H, m), 7.06-7.11 (2H, m), 7.28 (1H, d, *J* = 7.5 Hz), 7.57 (1H, d, *J* = 7.5 Hz), 7.64 (1H, br s).

4h: ¹H NMR (CDCl₃) δ: 0.69 (3H, s), 0.83 (3H, s), 0.87 (3H, d, *J* = 6.5 Hz), 0.87 (3H, d, *J* = 7 Hz), 0.92 (3H, d, *J* = 6.5 Hz), 0.69-1.65 (25H, m), 1.84-1.86 (1H, m), 2.00-2.04 (1H, m), 2.67-2.78 (2H, m), 6.99 (1H, t, *J* = 8 Hz), 7.04 (1H, t, *J* = 8 Hz), 7.25 (1H, d, *J* = 8 Hz), 7.67 (1H, br s), 7.70 (1H, d, *J* = 8 Hz). ¹³C NMR (CDCl₃) δ: 12.1, 12.2, 18.7, 20.6, 21.8, 22.6, 22.8, 23.9, 24.2, 25.0, 28.0, 28.3, 31.9, 34.1, 35.2, 35.8, 36.2, 37.0, 39.5, 40.2, 42.6, 46.1, 52.7, 56.3, 56.6, 110.4, 112.3, 118.8, 119.9, 120.2, 128.0, 132.9, 136.2.

- (1) Stoermer, D.; Heathcock, C. H. *J. Org. Chem.* **1993**, *58*, 564-568.
- (2) Wender, P. A.; Cooper, C. B. *Tetrahedron* **1986**, *42*, 2985-2991.
- (3) Naruse, Y.; Ito, Y.; Inagaki, S. *J. Org. Chem.* **1991**, *56*, 2256-2258.
- (4) Owellen, R. J. *J. Org. Chem.* **1974**, *39*, 69-72.

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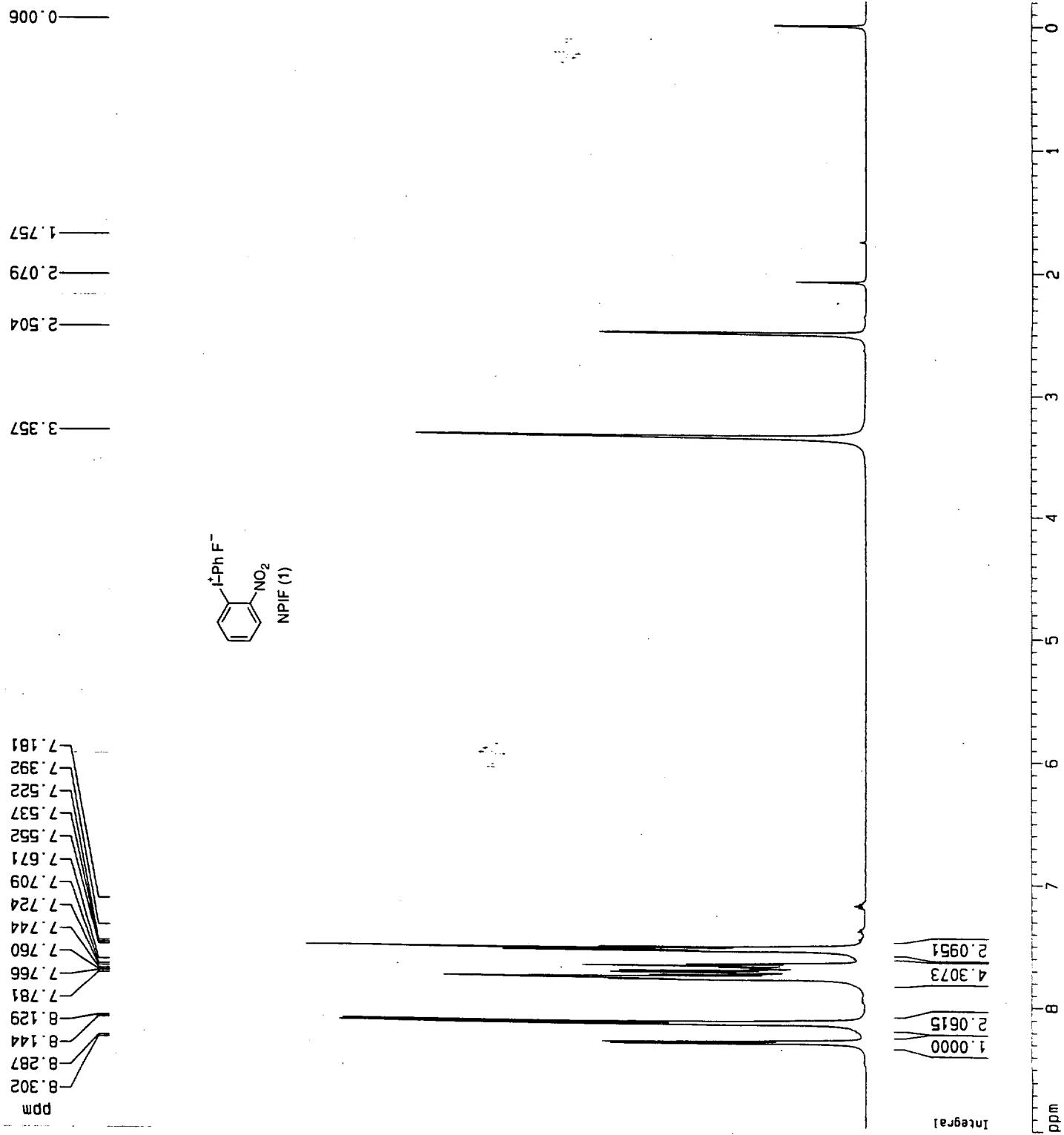
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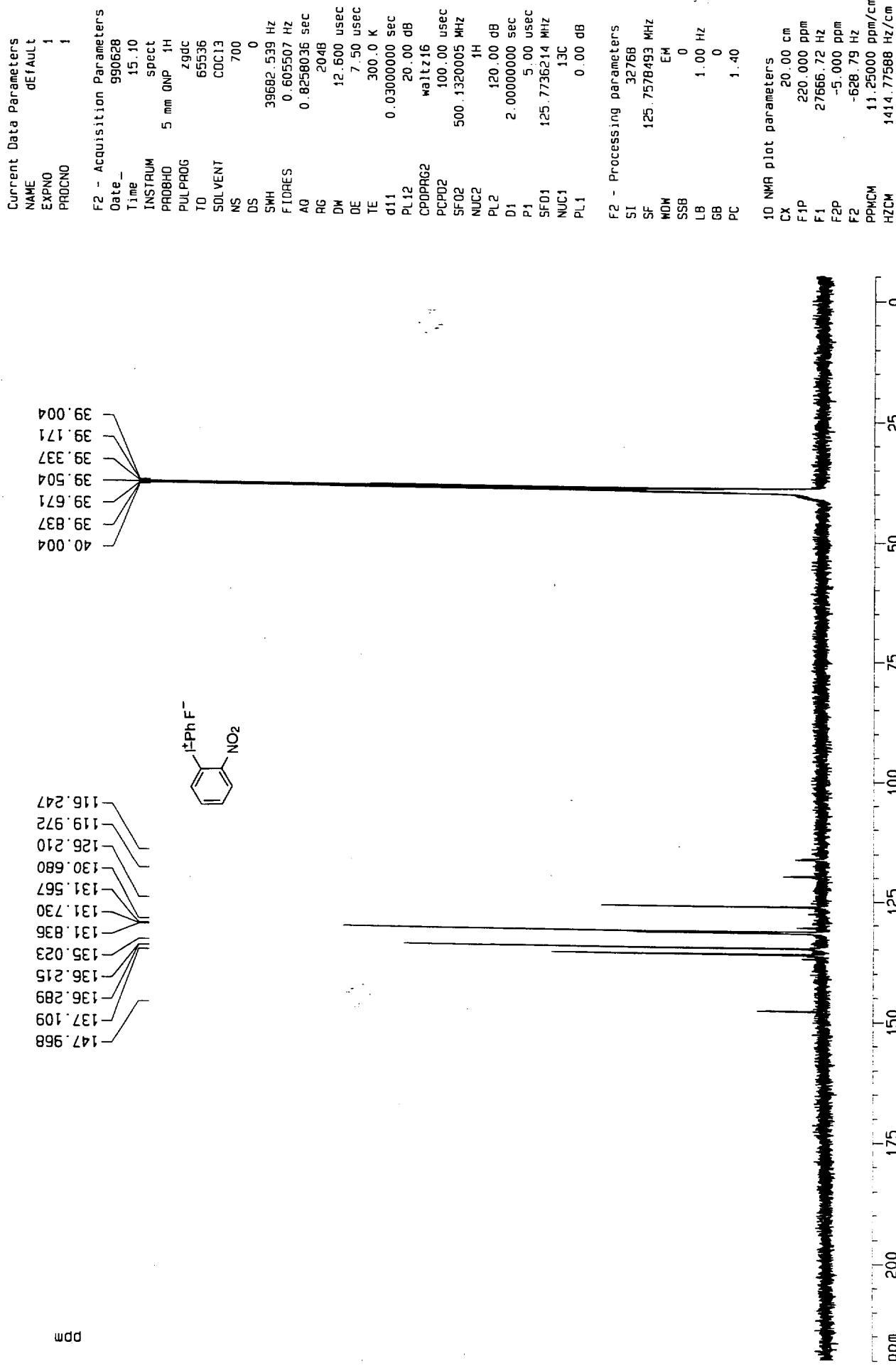
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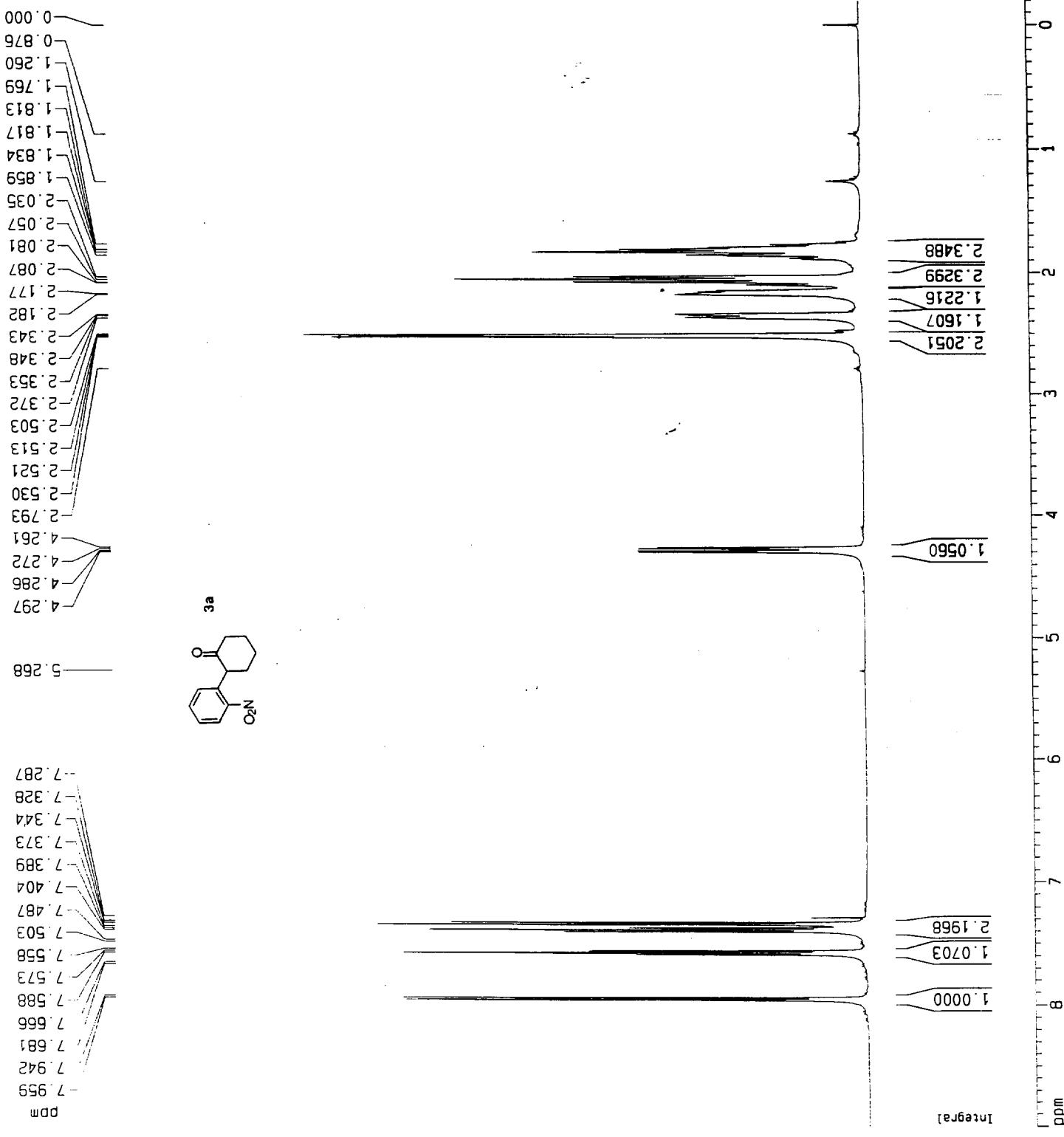
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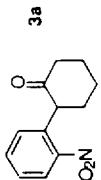
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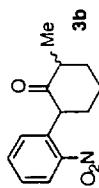
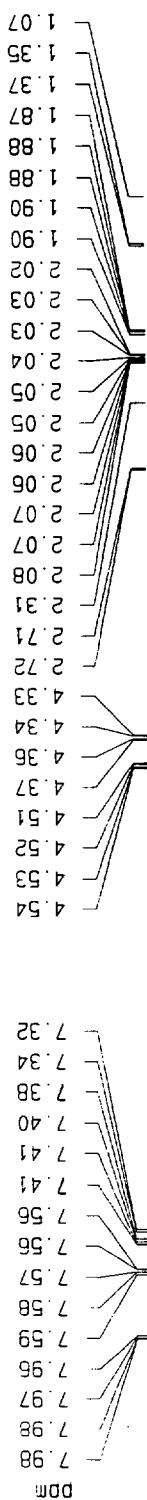
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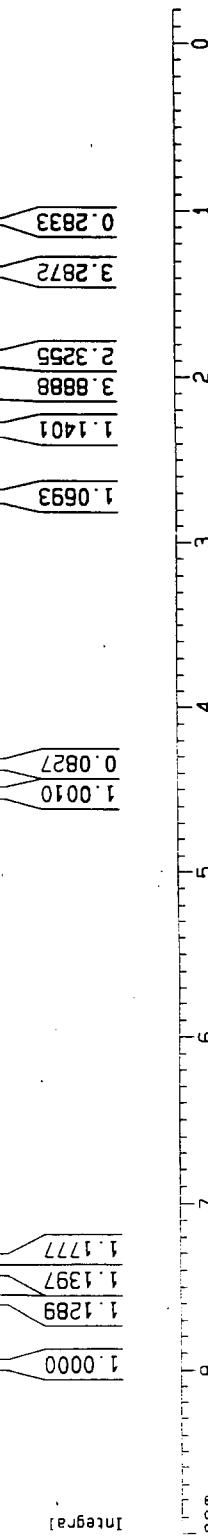
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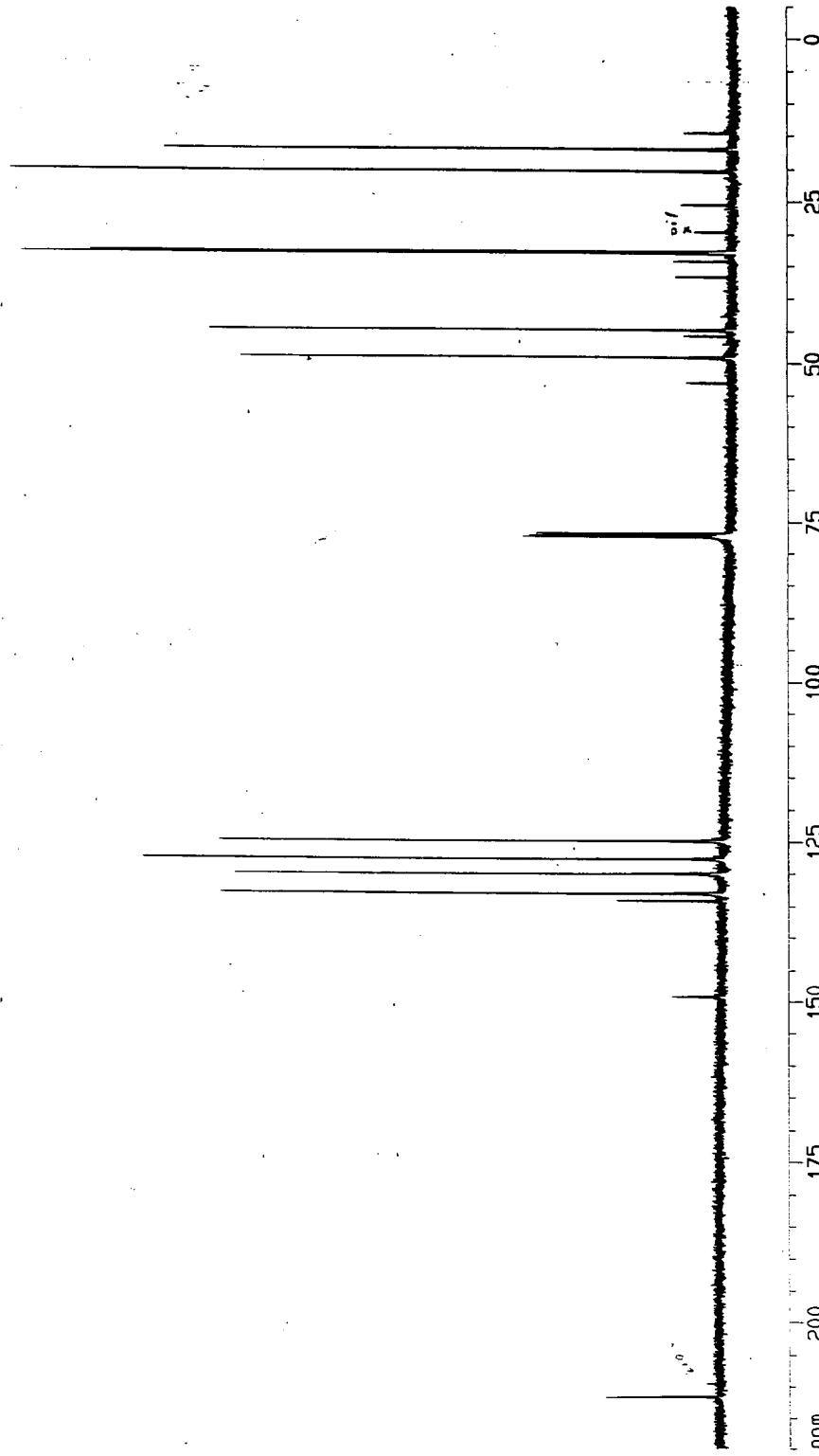
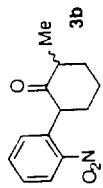
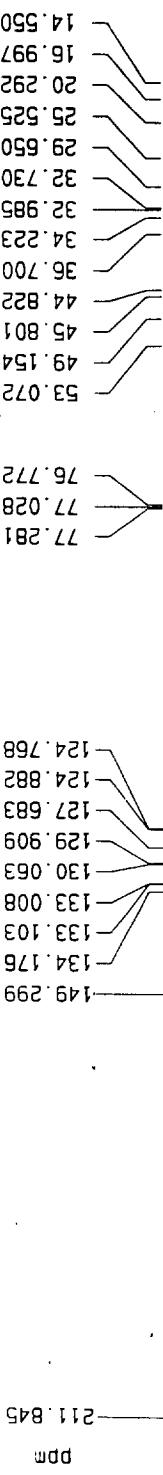


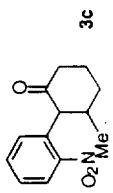
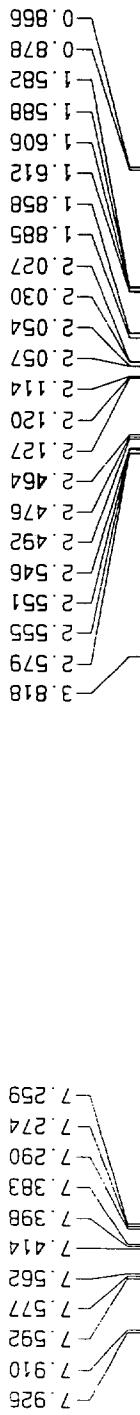
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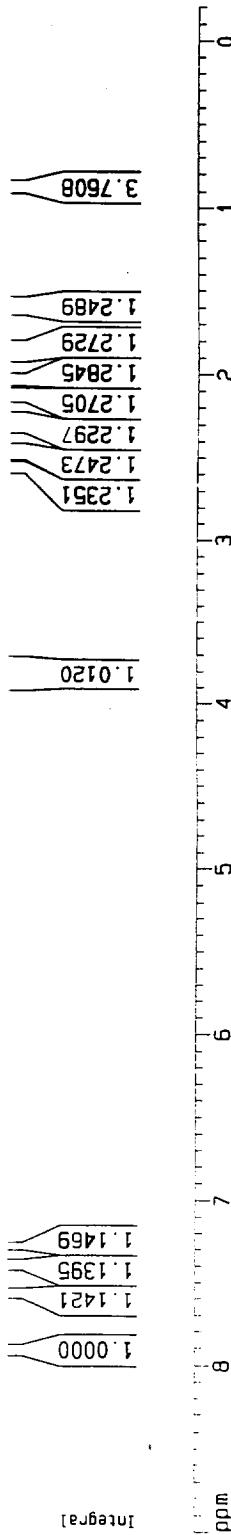


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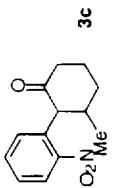
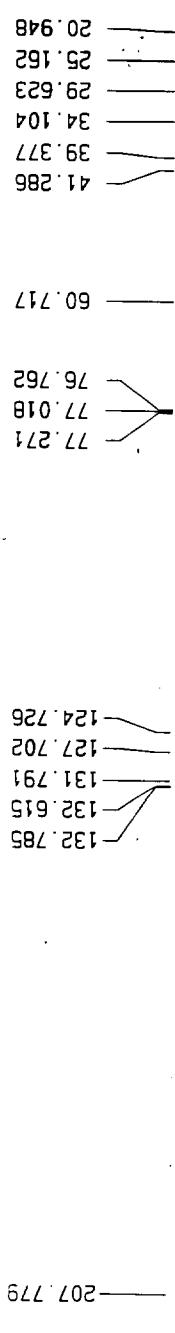


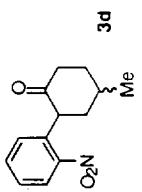
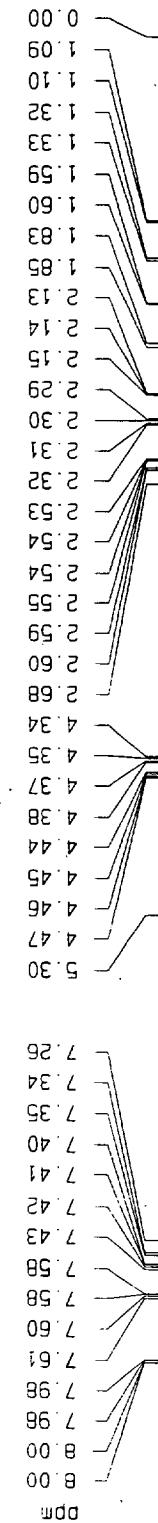
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Time 20.22
INSTRUM spect
PROBHD 5 mm QNP 1H
PULPROG 2gdc
TD 65536
SOLVENT CDCl3
NS 300
DS 0
SWH 39662.539 Hz
FIDRES 0.605507 Hz
AQ 0.8958036 sec
RG 1024
DW 12.600 usec
DE 7.50 usec
TE 300.0 K
d1 0.0300000 sec
PL12 wait216
CPDPG2 100.00 usec
PCPD2 500.1320005 MHz
SF02 NUC2
NUC2 1H
PL2 120.00 dB
D1 2.0000000 sec
P1 5.00 usec
SF01 125.7736214 MHz
NUC1 13C
PL1 0.00 dB

F2 - Processing parameters
SI 32768
SF 125.7578007 MHz
MDW EH
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

1D NMR plot parameters
CX 20.00 cm
F1P 220.000 ppm
F1 27656.71 Hz
F2P -5.000 ppm
F2 -628.79 Hz
PPMCM 11.25000 ppm/cm
HZCM 1414.77527 Hz/cm





F2 - Acquisition Parameters

Date_	990529
Time	12.47
INSTRUM	spect
PROBHD	5 mm GNP 1H
PULPROG	zg
TD	32768
SOLVENT	CDCl ₃
NS	8
DS	0
SWH	5208.333 Hz
FIRES	0.158946 Hz
AQ	3.1457779 sec
RG	128
DW	96.000 usec
DE	4.50 usec
TE	300.0 K
D1	1.0000000 sec
P1	5.00 usec
SFO1	500.1320118 MHz
NUC1	1H
PL1	0.00 dB

F2 - Processing parameters

SI	16384
SF	500.1300116 MHz
WDW	EM
SSB	0
LB	0.30 Hz
GB	0
PC	1.00

1D NMR plot parameters

CX	20.00 cm
F1P	9.000 ppm
F1	4501.17 Hz
F2P	-0.200 ppm
F2	-100.03 Hz
PPMCM	0.46000 ppm/cm
HZCM	230.05980 Hz/cm



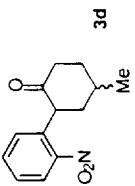
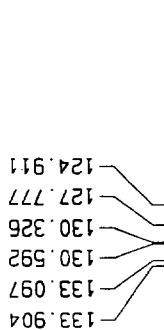
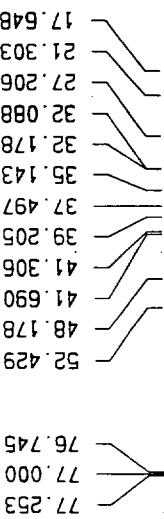
Current Data Parameters
 NAME dFAult 1
 EXPNO 1
 PROCN0

F2 - Acquisition Parameters

Date_ 990529
 Time 11.40
 INSTRUM spect
 PROBHD 5 mm QNP 1H
 PULPROG zgdc
 TD 65536
 SOLVENT CDCl3
 NS 512
 DS 0
 SWH 39682.539 Hz
 FIDRES 0.605507 Hz
 AQ 0.8258036 sec
 RG 1024
 DW 12.600 usec
 DE 7.50 usec
 TE 300.0 K
 d11 0.03000000 sec
 PL12 20.00 dB
 CPDPRG2 Waltz16
 PCPD02 100.00 usec
 SF02 500.1320005 MHz
 NUC2 1H
 PL2 120.00 dB
 D1 2.0000000 sec
 P1 5.00 usec
 SF01 125.7736214 MHz
 NUC1 13C
 PL1 0.00 dB

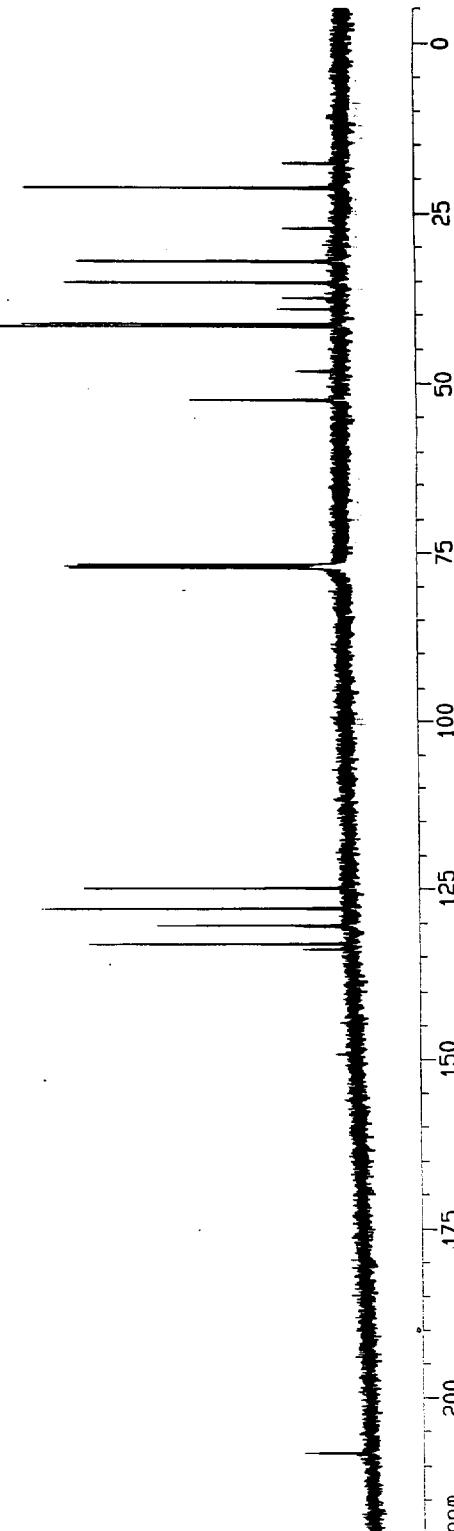
F2 - Processing parameters

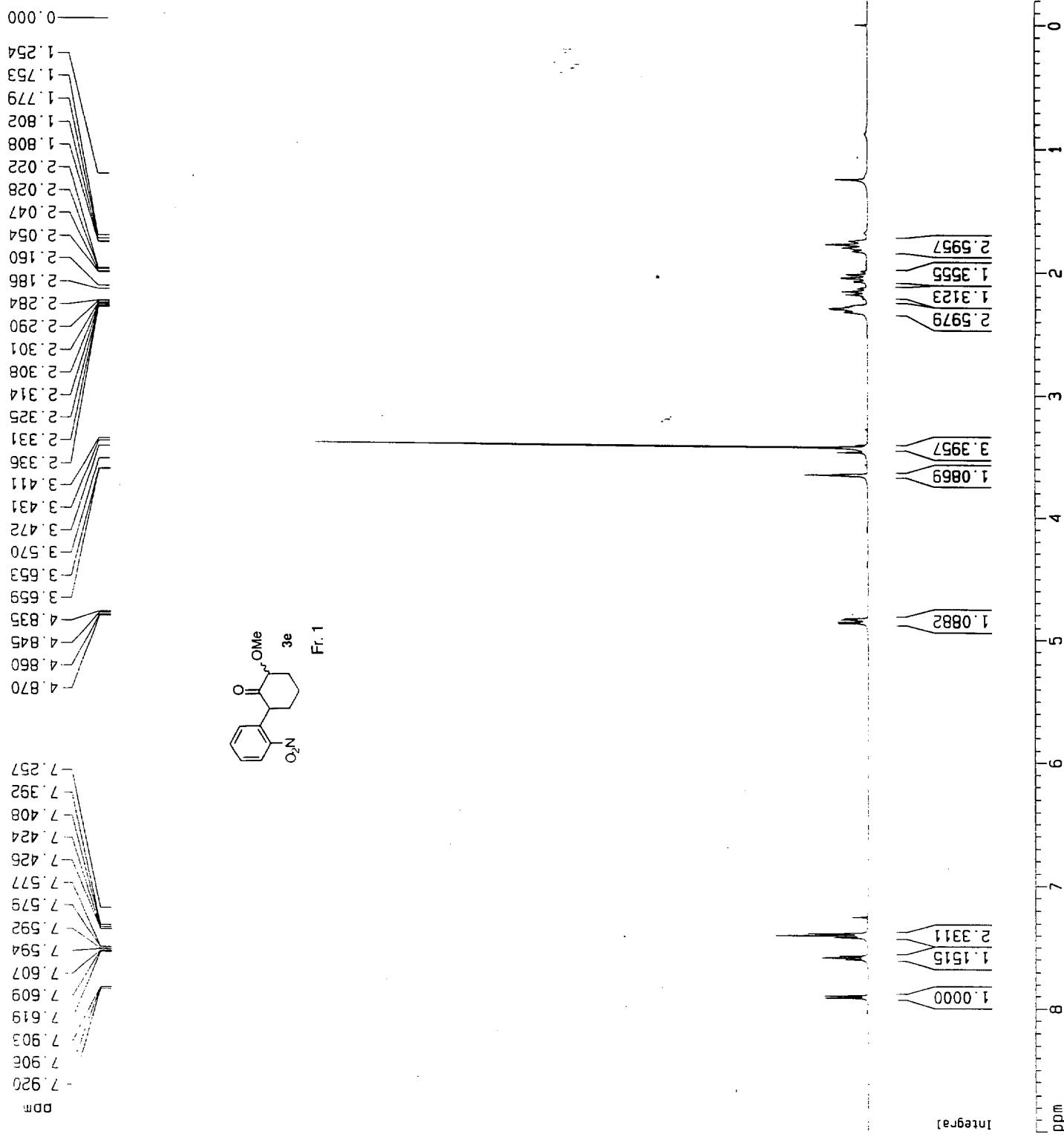
SI 32768
 SF 125.7577955 MHz
 MDW EM
 SS8 0
 LB 1.00 Hz
 F2P -5.00 ppm
 F2 -628.79 Hz
 PPMCM 11.25000 ppm/cm
 HZCM 1414.77527 Hz/cm



208.274

ppm



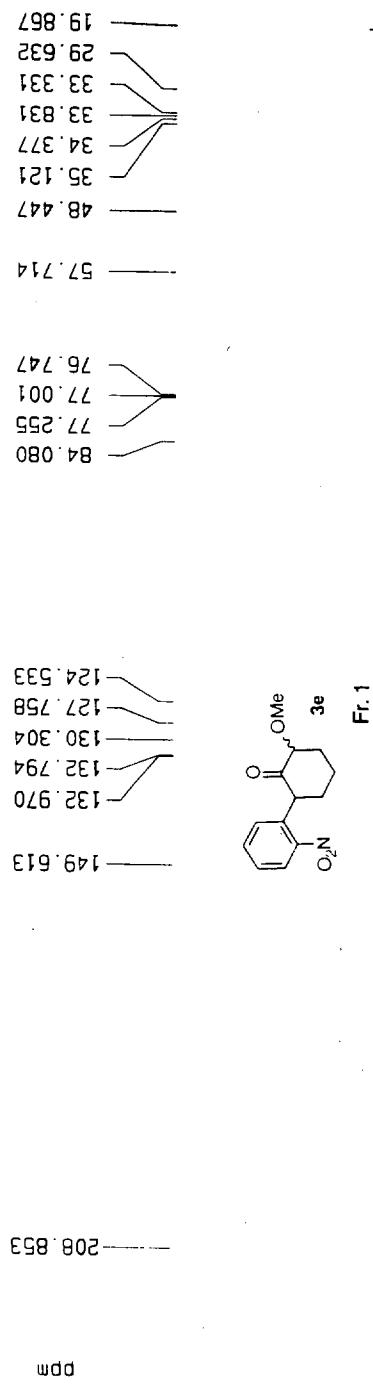


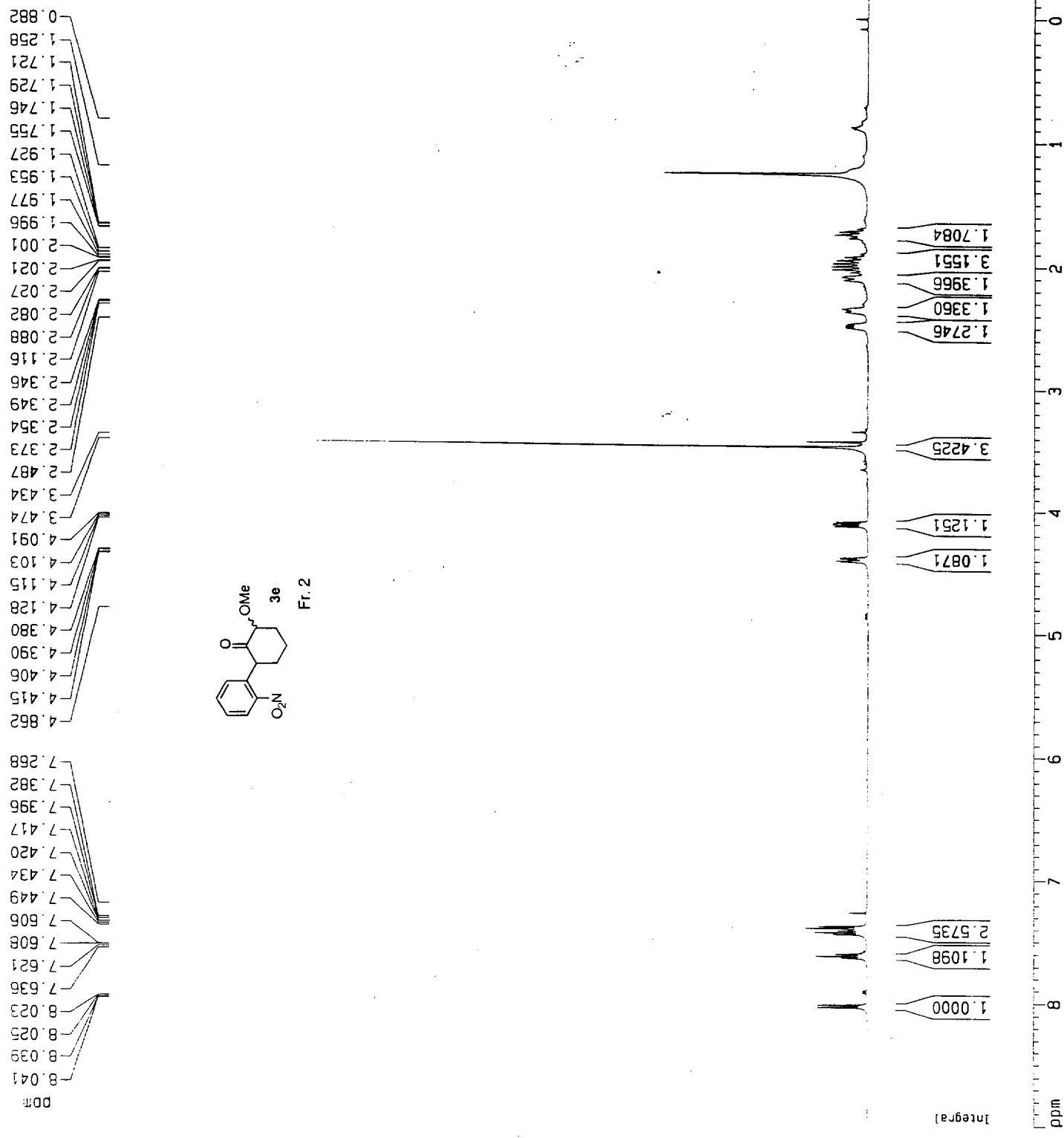
Current Data Parameter:
NAME dEFAULT
EXPNO
PROCND

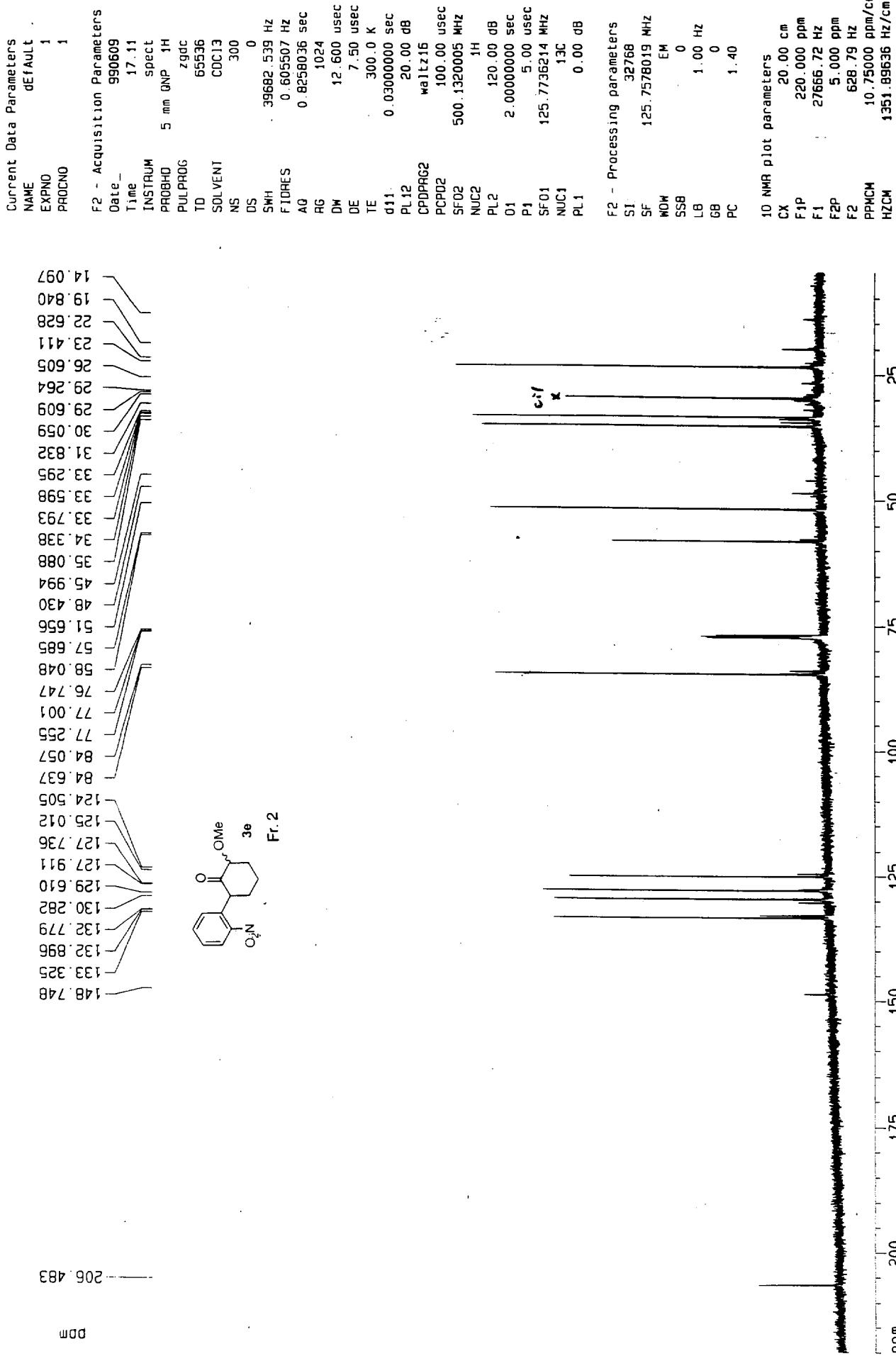
F2 - Acquisition Parameter
Date 990501
Time 16:41
INSTRUM PROBHD
PULPROG 5 mm QNP 1H
TD 65536
SOLVENT CDCl₃
NS 301
DS 39682.53!
SWH 0.60550.
FIDRES 0.025603F
RG 20.4F
DW 12.60!
DE 7.51
TE 300.1
d1 0.03000001
PL12 20.01
CPDPRG2 Waltz11
PCPDQ2 100.01
SF02 500.132000!
NUC2 11
PL2 120.01
D1 2.00000001
P1 5.01
SF01 125.773621.
NUC1 13C
PL1 0.01

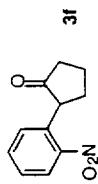
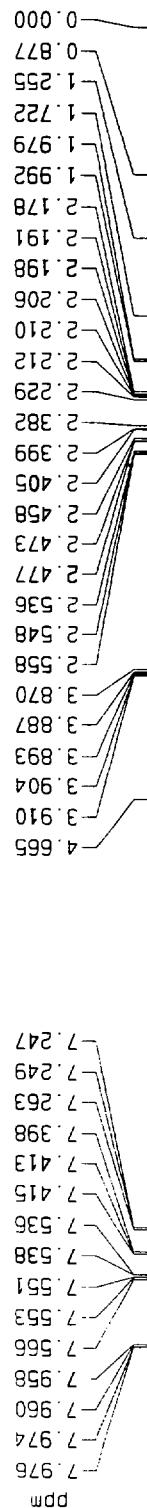
F2 - Processing parameters
SI 3278
SF 125.757799.
MW 1
SSB 1.01
LB 1
GB 1
PC 1.41

1D NMR plot parameters
CX 20.01
F1P 230.00C
F1 28924.25
F2P -5.00C
F2 -629.75
PHPCM 11.7500C
HZCW 1477.6540C









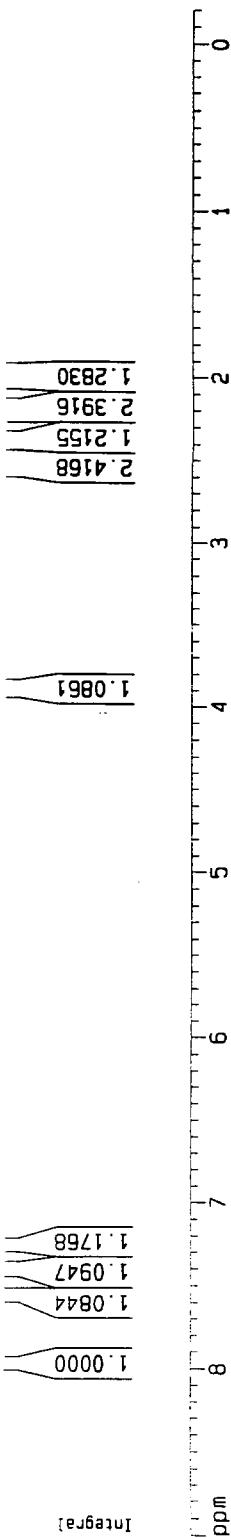
Current Data Parameters
NAME default
EXPNO 1
PROCNO 1

F2 - Acquisition Parameters
Date 990608
Time 20.40
INSTRUM spect
PROBHD 5 mm DNP 1H
PULPROG zg32768
TD 32768
SOLVENT CDCl3
NS 8
DS 0
SWH 5208.333 Hz
FIDRES 0.158946 Hz
AQ 3.1457779 sec
RG 32
DW 96.000 usec
DE 4.50 usec
TE 300.0 K
d11 0.0300000 sec
PL12 120.00 dB
CPDPG2

PCPD2 100.00 usec
SF02 500.1300000 MHz
NUC2 off
PL2 120.00 dB
D1 1.0000000 sec
P1 5.00 usec
SF01 500.1320118 MHz
NUC1 1H
PL1 0.00 dB

F2 - Processing parameters
SI 16384
SF 500.1300085 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

1D NMR plot parameters
CX 20.00 cm
F1P 9.000 ppm
F1 4501.17 Hz
F2P -0.200 ppm
F2 -100.03 Hz
PPMCM 0.06000 ppm/cm
HZCM 230.05980 Hz/cm

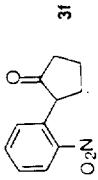
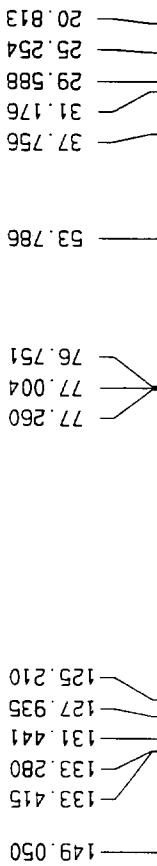


Current Data Parameter
dFAul
NAME EXPNO
PROCNO

F2 - Acquisition Param
Date 99060
Time 20.5
INSTRUM spec
PROBHD 5 mm QNP 1
PULPROG zgd
TD 6553
SOLVENT C6C1
NS 30.
DS 39682.53
SWH 0.60550
FTDRES 0.825803
AQ 0.03000001
RG 102.
DW 12.60
DE 7.51
TE 300.
d11 0.03000001
PL12 20.01
CPDPBG2 Waltz11
PCPD2 100.01
SF02 500.13200001
NUC2 11
PL2 120.01
D1 2.00000001
P1 5.01
SF01 125.773621
NUC1 131
PL1 0.01
PL1

F2 - Processing param
SI 3276f
SF 125.757805f
WDW Et
SSB 1.0f
LB 28924.2f
GB 1.4f
PC

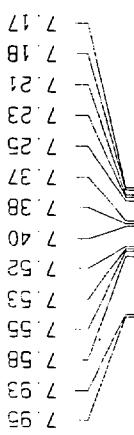
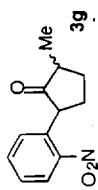
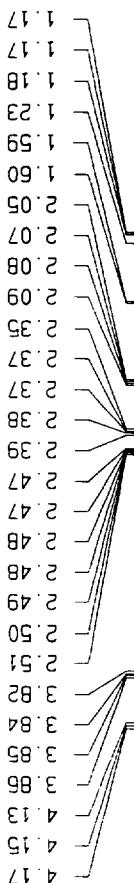
1D NMR plot parameters
CX 20.0f
F1P 230.00f
F1 28924.2f
F2P -5.00f
F2 -628.7f
PPM 11.7500f
HZCM 1477.6540f



215.391

ppm





DDM

Current Data Parameters
NAME NAME default
EXPNO EXPNO 1
PROCNO PROCNO 1

F2 - Acquisition Parameters

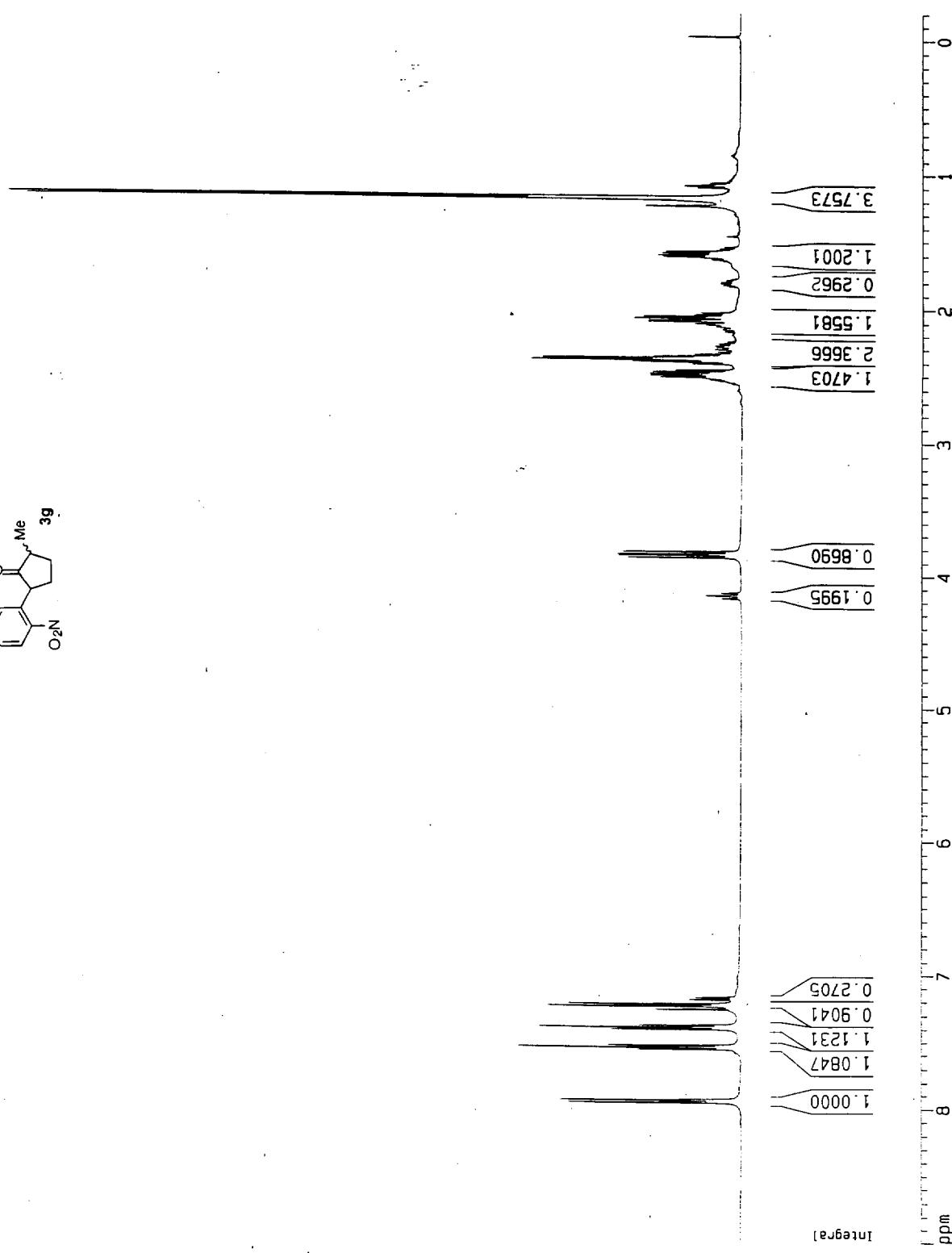
Date_ 990608
Time 18.19
INSTRUM spect
PROBHD 5 mm QNP 1H
PULPROG zg
TD 32768
SOLVENT CDCl3
NS 8
DS 0
SWH 5208.333 Hz
FIDRES 0.158946 Hz
AQ 3.1457779 sec
RG 32
DW 96.000 usec
DE 4.50 usec
TE 300.0 K
D1 1.0000000 sec
P1 5.00 usec
SF01 500.132018 MHz
NUC1 1H
PL1 0.00 dB

F2 - Processing parameters

SI 16384
SF 500.1300181 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

1D NMR plot parameters

CX 20.00 cm
F1P 9.000 ppm
F1 4501.17 Hz
F2P -0.200 ppm
F2 -100.03 Hz
PPMC 0.46000 ppm/cm
HZCM 230.05980 Hz/cm



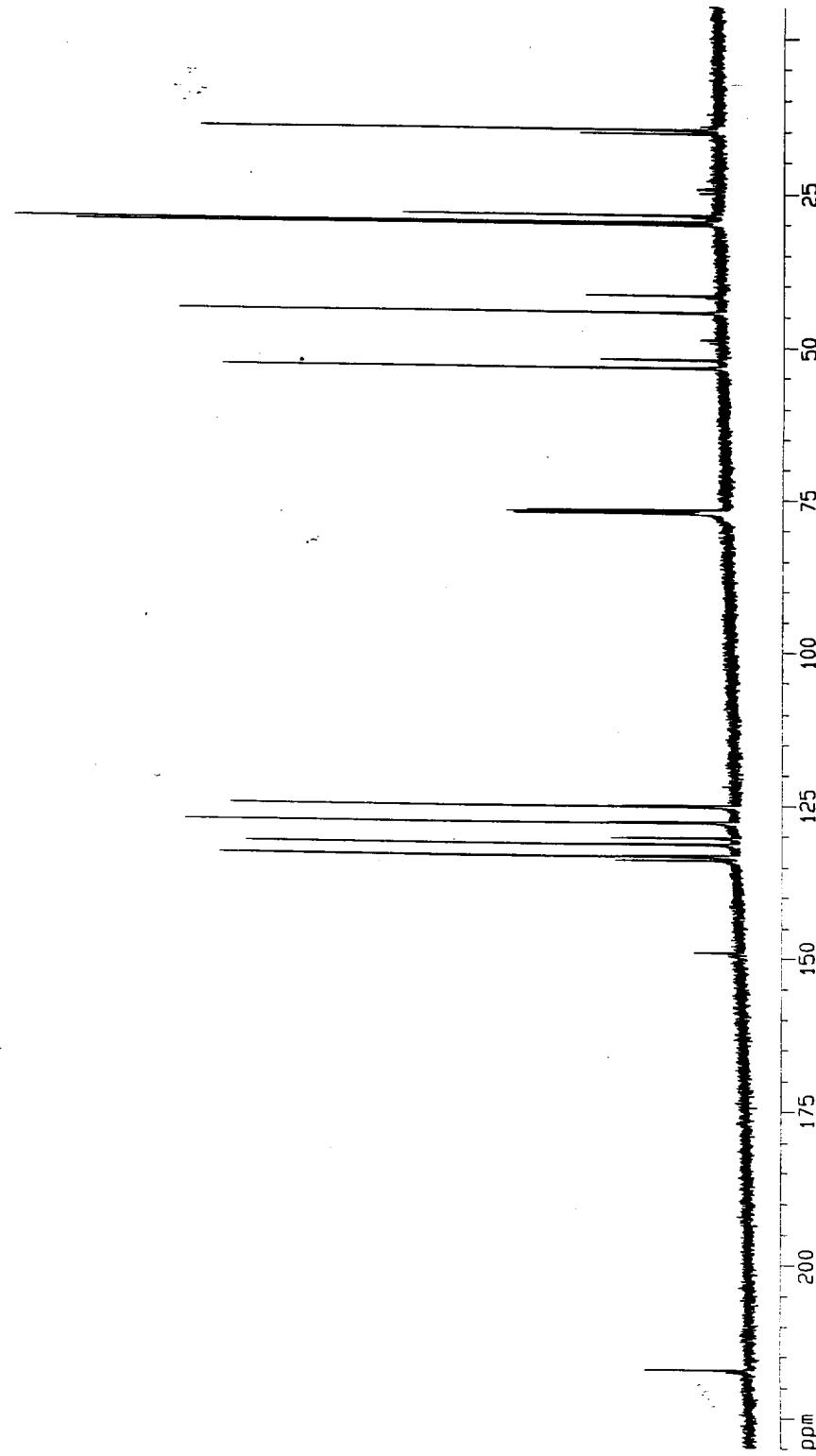
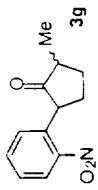
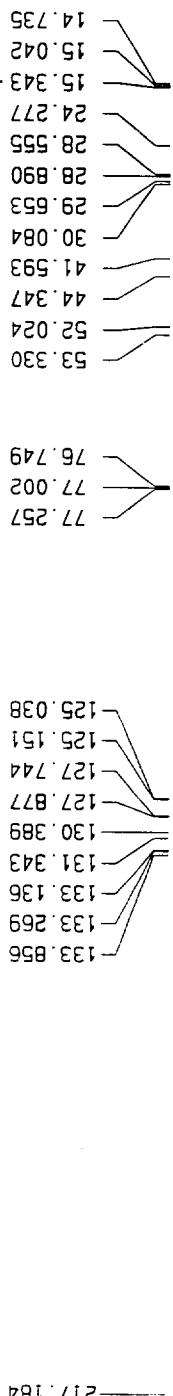
Integral

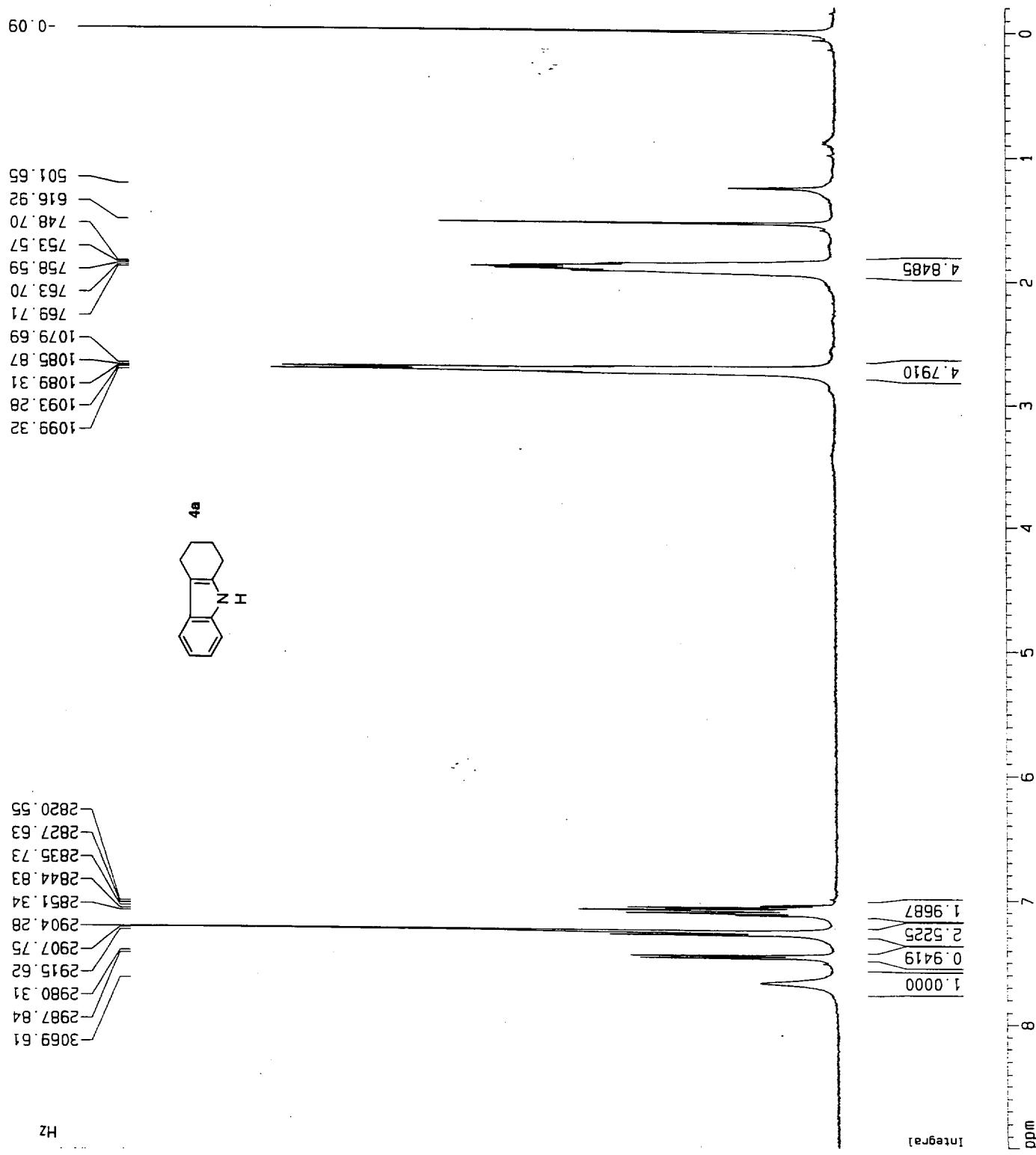
ppm

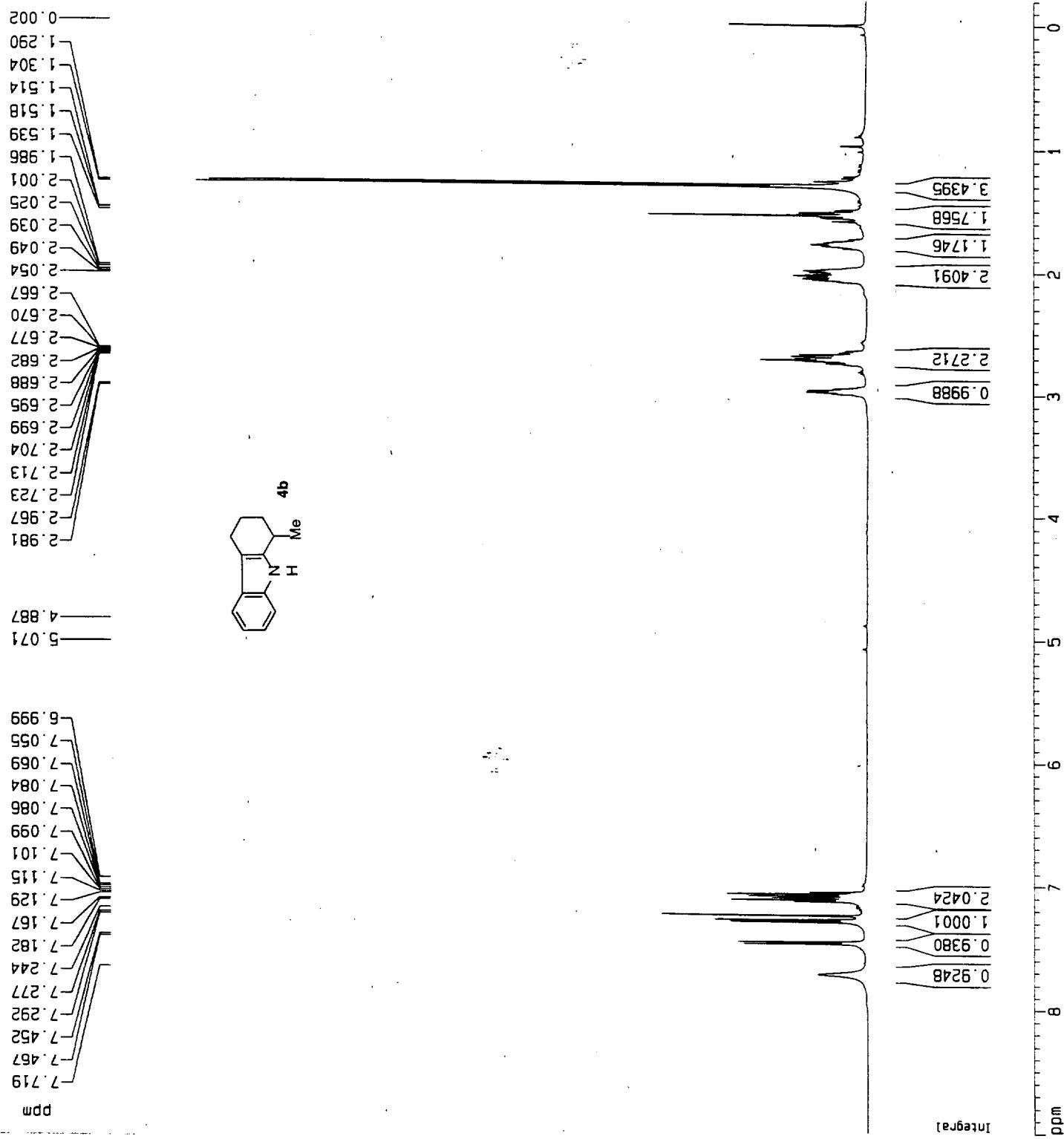
Current Data Parameters
NAME dEfAu
EXPNO
PROCNO

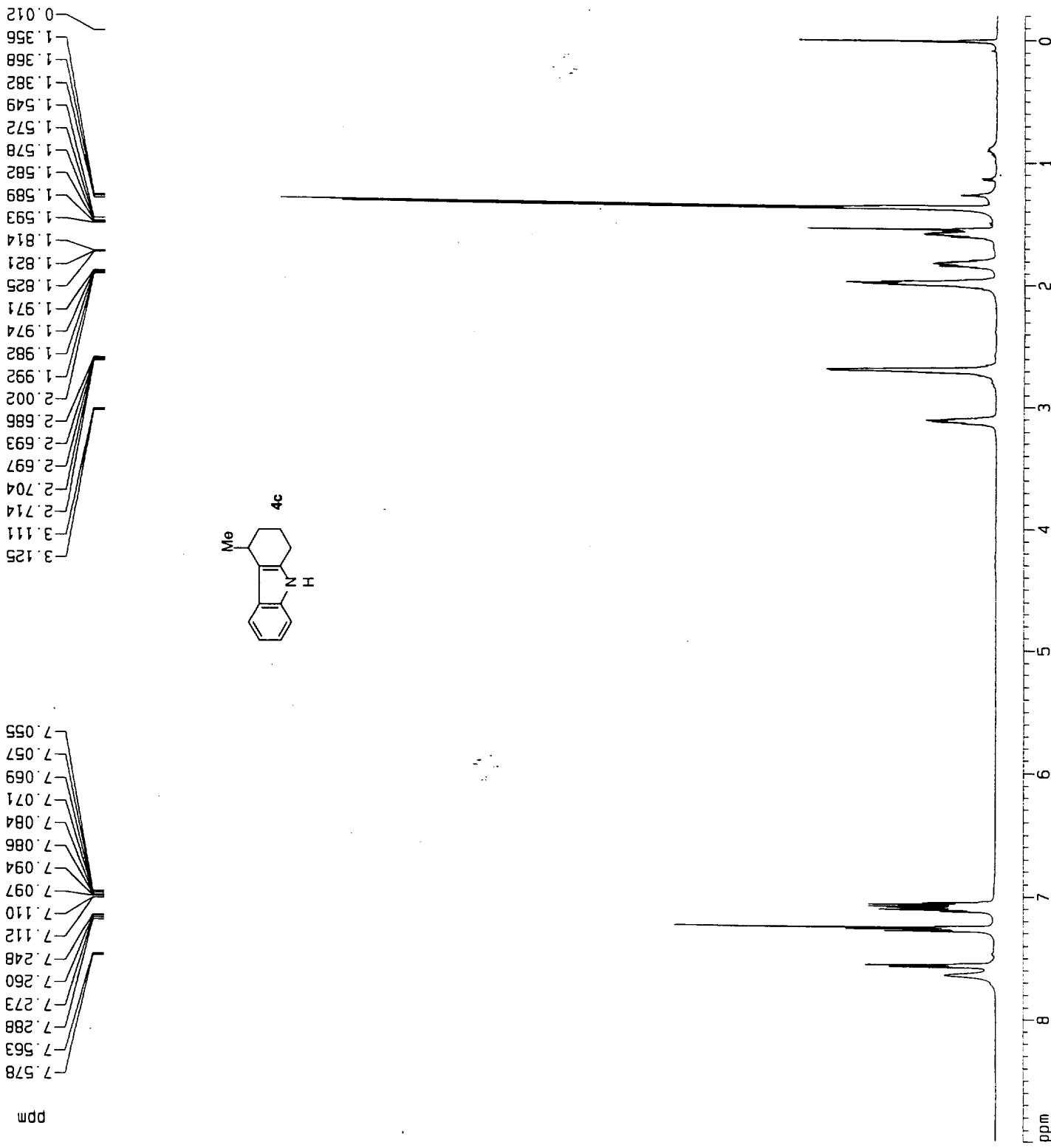
F2 - Acquisition Para
Date_ 9905
Time_ 18:
INSTRUM spe
PROBHD 5 mm QNP
PULPROG zg
TD 655
SOLVENT CDCl₃
NS 5
DS 39682.5
SWH 0.60550
FIDRES 0.82380
RG 10⁴
DW 12.66
DE 7.5
TE 300.
d11 0.0300000
PL12 20.0
CPDPG2 waltz1
PCPD2 100.0
SF02 500.132000
NUC2 1
PL2 120.0
P1 2.0000000
SF01 5.0
NUC1 13
PL1 0.0
PC 0.0

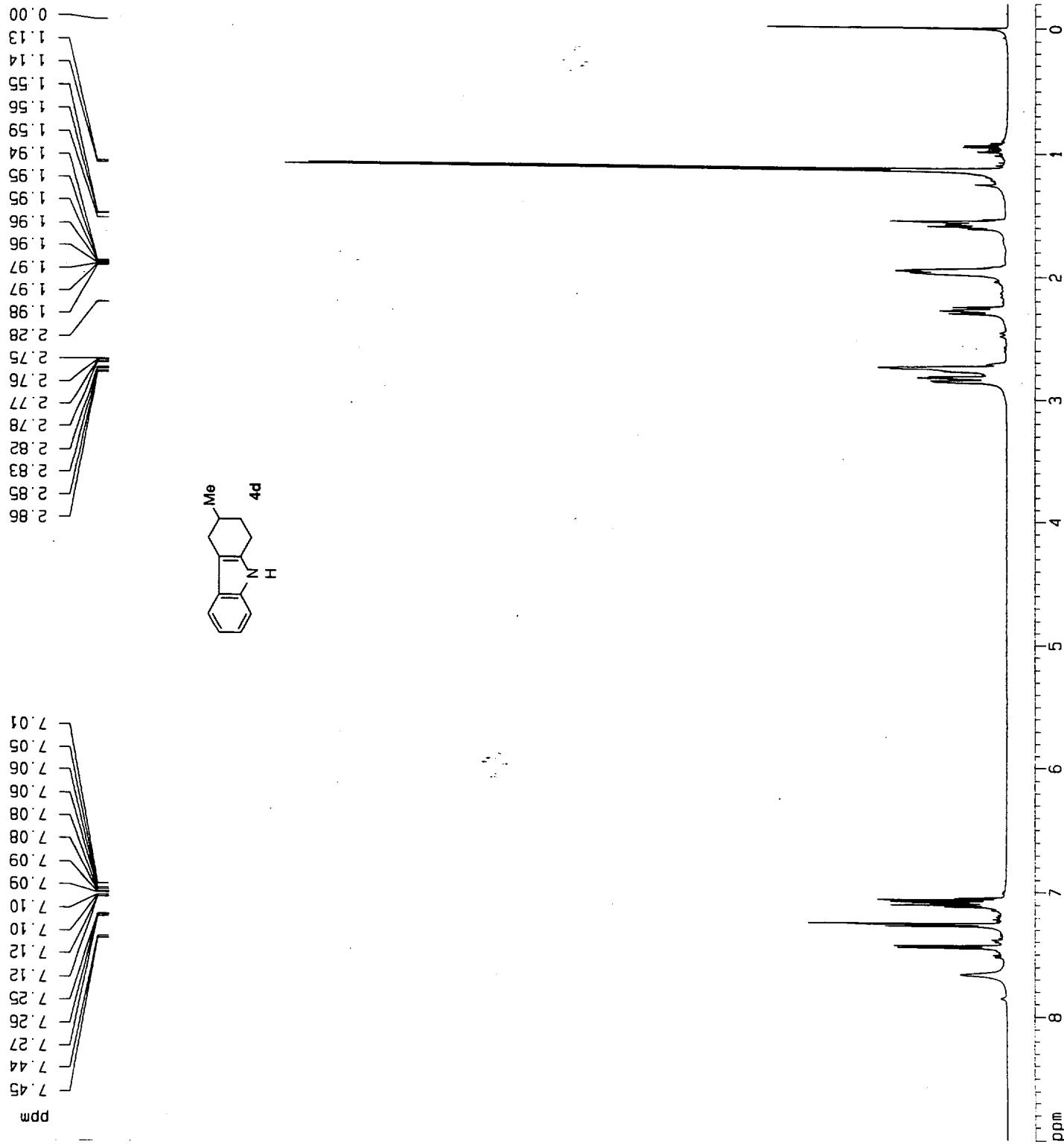
F2 - Processing para
SI 3276
SF 125.757801
NDW E_i
SSB 1.0
LB GB
PPMCM 11.75001
HZCM 1477.65401











Current Data Parameters
 NAME dEfault 1
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date 990605
 Time 13.25
 INSTRUM spect
 PROBHD 5 mm QNP 1H
 PULPROG zg
 TD 32768
 SOLVENT CDCl3
 NS 8
 DS 0
 SWH 4194.631 Hz
 FIDRES 0.128010 Hz
 A0 3.9059956 sec
 RG 161.3
 DW 119.200 usec
 DE 4.50 usec
 TE 300.0 K
 D1 1.0000000 sec

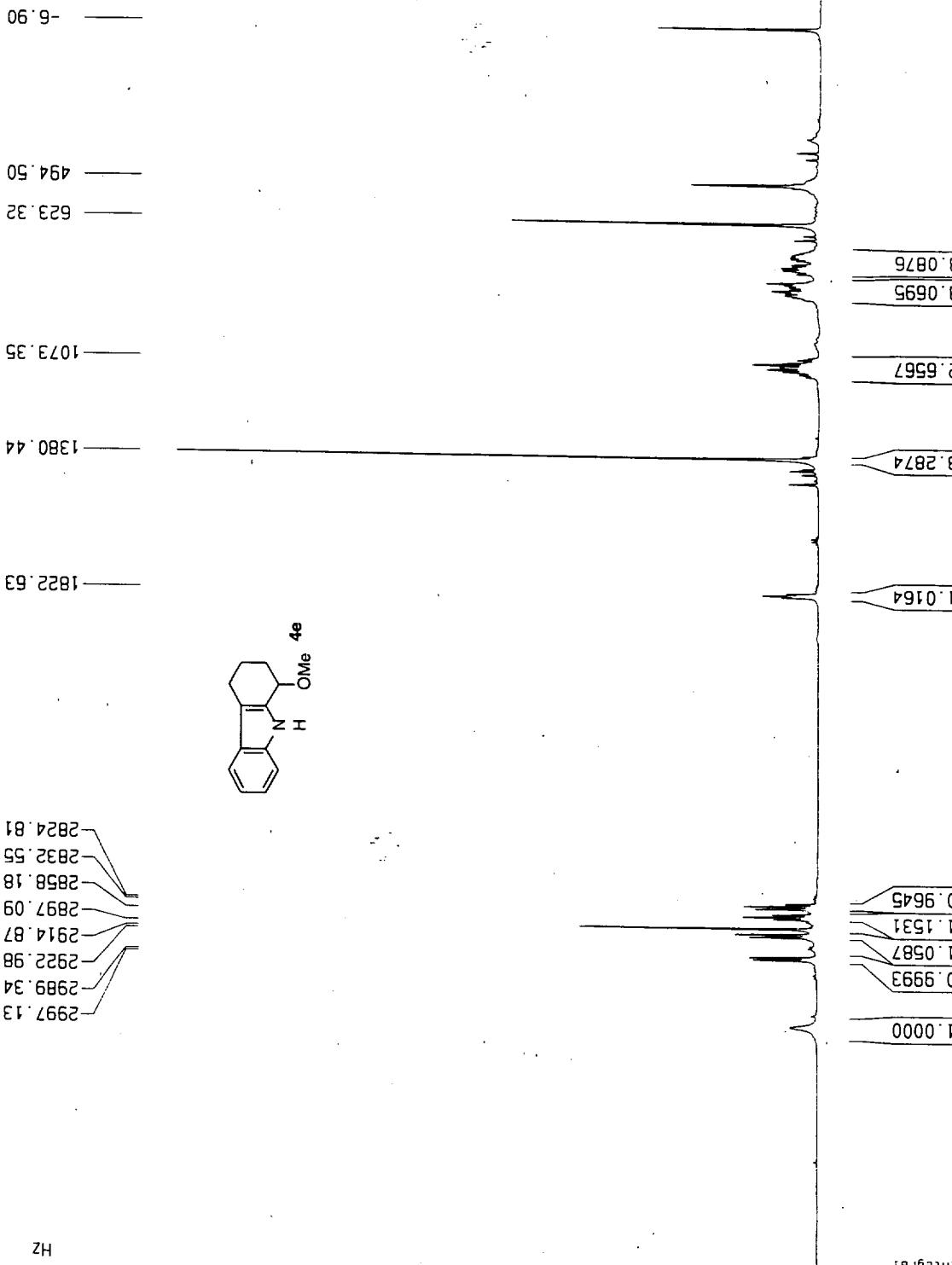
===== CHANNEL f1 =====

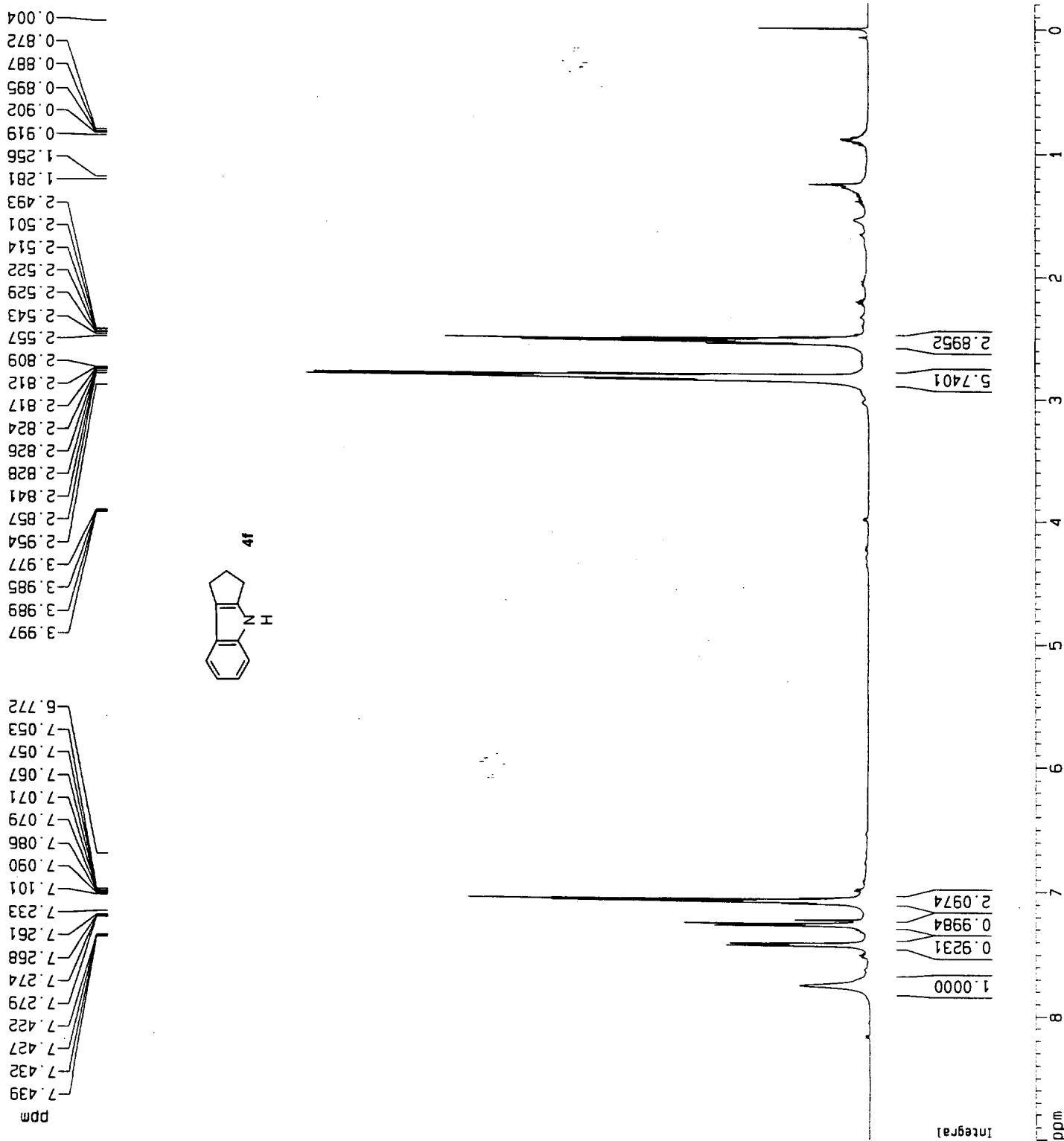
NUC1 1H
 P1 8.50 usec
 PL1 0.00 dB
 SF01 400.139246 MHz

F2 - Processing parameters

S1 16384
 SF 400.1300177 MHz
 WDM EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

1D NMR plot parameters
 CX 20.00 cm
 F1P 10.007 ppm
 F1 4004.18 Hz
 F2P -0.476 ppm
 F2 -190.45 Hz
 PPMCM 0.52/1.16 ppm/cm
 HZCM 209.73154 Hz/cm





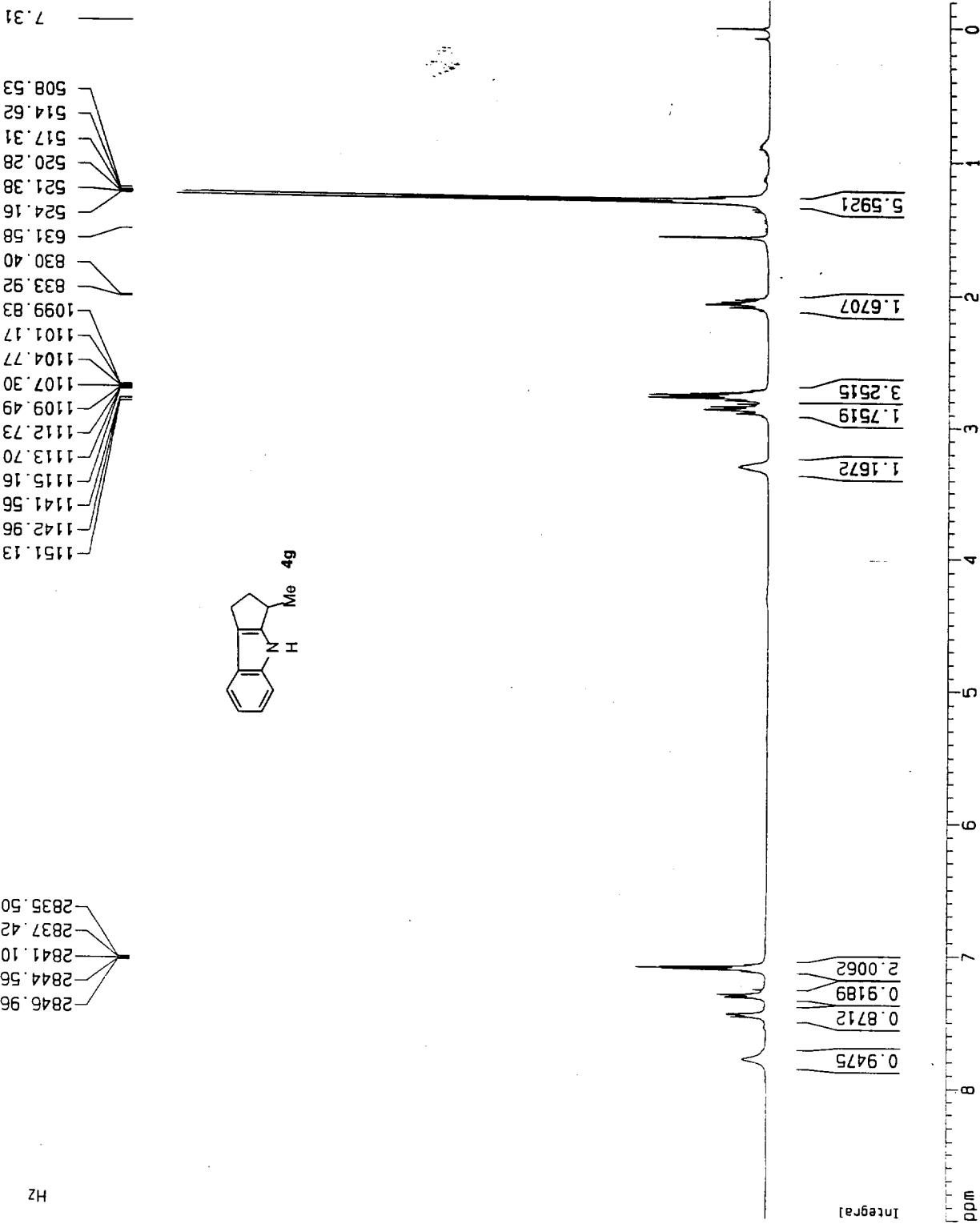
Current Data Parameters
 NAME DEFault
 EXPNO 1
 PROCNO 1

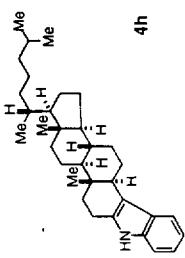
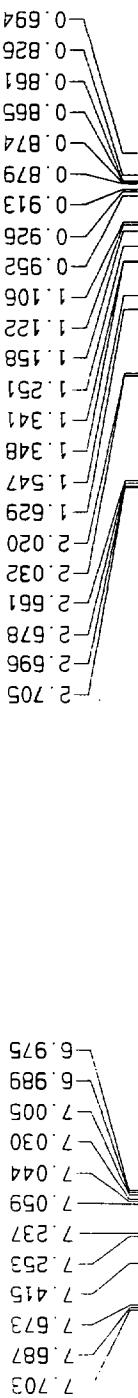
F2 - Acquisition Parameters
 Date 990603
 Time 17.34
 INSTRUM spect
 PROBHD 5 mm QNP 1H
 PULPROG TD
 32768
 SOLVENT CDCl3
 NS 8
 DS 0
 SWH 4194.631 Hz
 FIDRES 0.128010 Hz
 AQ 3.9059956 sec
 RG 101.6
 DW 119.200 usec
 DE 4.50 usec
 TE 300.0 K
 D1 1.0000000 sec

===== CHANNEL f1 =====
 NUC1 1H
 P1 8.50 usec
 PL1 0.00 dB
 SF01 400.1319246 MHz

F2 - Processing parameters
 SI 16384
 SF 400.1300072 MHz
 MW 0
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

1D NMR plot parameters
 CX 20.00 cm
 F1P 9,000 ppm
 F1 3601.17 Hz
 F2P -0.200 ppm
 F2 -80.03 Hz
 PPMLW 0.46000 ppm/cm
 HZCM 184.05980 Hz/cm



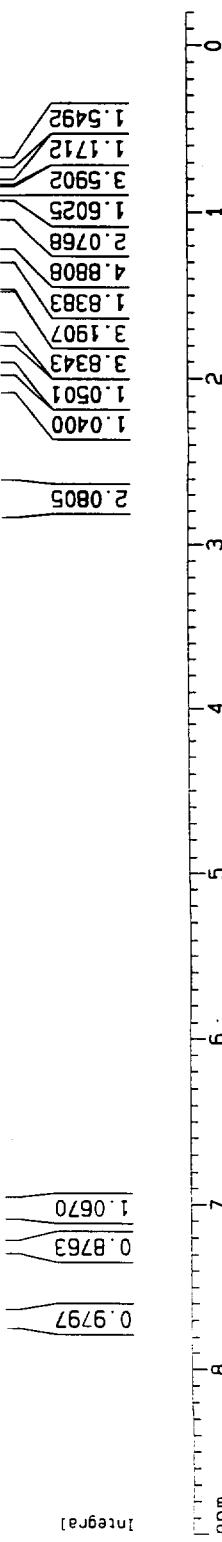


Current Data Parameters
NAME dEfault
EXPNO 1
PROCNO 1

F2 - Acquisition Parameters
Date_ 990612
Time 17.30
INSTRUM spect
PROBID 5 mm QNP 1H
PULPROG 32768
TD 32768
SDLVENT CDCl₃
NS 8
DS 0
SWH 5208.333 Hz
FIDRES 0.158946 Hz
AQ 3.145779 sec
RG 128
DW 96.000 usec
DE 4.50 usec
TE 300.0 K
d11 0.0300000 sec
PL12 120.00 dB
CPDPG2
PCPD2 100.00 usec
SF02 500.1300000 MHz
NUC2 off
PL2 120.00 dB
D1 1.0000000 sec
P1 5.00 usec
SF01 500.1320118 MHz
NUC1 1H
PL1 0.00 dB
PC 1.00

F2 - Processing parameters
SI 16384
SF 500.1300212 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

1D NMR pilot parameters
CX 20.00 cm
F1P 9.000 ppm
F1 4501.17 Hz
F2P -0.200 ppm
F2 -100.03 Hz
PPNCH 0.46000 ppm/cm
HZCM 230.05981 Hz/cm



Current Data Parameters

NAME	dEfault
EXPNO	1
PROCNO	1

F2 - Acquisition Parameters

Date	990612
Time	19:08
INSTRUM	spect
PROBHD	5 mm QNP 1H
PULPROG	2gdc
TD	65536
SOLVENT	CDC13
NS	2024
DS	0
SWH	39682.539 Hz
FIDRES	0.605507 Hz
AQ	0.8258036 sec
RG	2048
DW	12.600 usec
DE	7.50 usec
TE	300.0 K
d111	0.03000000 sec
PL12	20.00 dB
CPDPFG2	Wait16
PCPD2	100.00 usec
SFO2	500.1320005 MHz
NUC2	1H
PL2	120.00 dB
D1	2.00000000 sec
P1	5.00 usec
SFO1	125.7736214 MHz
NUC1	13C
PL1	0.00 dB

F2 - Processing parameters

CX	20.00 cm
F1P	220.000 ppm
SI	16384
SF	125.7577934 MHz
NDW	EM
SSB	0
LB	0.30 Hz
GB	0
PC	1.40
PPMCM	11.25000 ppm/cm
HZCM	1414.77515 Hz/cm

ppm

136.187
132.879
120.220
119.889
118.826
112.267
110.417
77.255
76.752
56.585
56.317
52.698
46.128
40.197
35.207
36.175
39.520
34.149
31.876
28.313
25.001
24.209
23.864
22.824
21.824
20.648
18.665
12.193
12.116

